Product Code: 1006

# HLA-B\*27 Single Box 1.0 Typing Kit

In vitro diagnostics disposal

# **Instructions Manual**



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#### Presentation

HLA-B\*27 has been associated with several autoimmune and infection diseases, such as: rheumatoid arthritis, ankylosing spondylitis, spondyloarthropathies, uveitis, Crohn's and Bechterew's disease. HLA-B\*27 allele information is very helpful to clarify association with disease. With GENEBOX HLA-B\*27 TYPING KIT is possible to identify the presence or absence of HLA-B\*27.

This kit contains typing plates with dried primer mixes and PCR Master Mix for HLA-B\*27 detection.

# Product Changes and Improvements

The HLA-B\*27 Single Box specificity and interpretation tables are constantly updated, to include new associations described. This product can also be improved in order to increase the yield of the specific PCR product.

The primers exchanged, added or modified, compared to the previous lot, are detailed in the table below.

Tube	primers	motivation
N/A		

# **Quality Control**

The following DNA samples from the 13th International Histocompatibility Workshop SSOP Panel were used to verify the kit primers mixes specificity.

Workshop Name	Designation
IHW 09266	PAR
IHW 09376	FH4
IHW 09377	FH5
IHW 09380	FH6

No false positive or negative amplifications were obtained.

The negative control tube can detect cross-contamination with PCR products.

#### Cell line validation sheet

	HLA-B*27 plus SSP typing kit				
Cel	l line	Cell Typing			Kit positive wells
00.	0	HLA-A*	HLA-B*	HLA-Cw*	The positive world
9380	FH6	2402;2901	2702;0705/6	02022;1505	All
9376	FH4	0101	2703;2705	02022	All
9266	PAR	11011;2402	2706;4801	03041;0801	All
9377	FH5	2902;0201	2709;4403	0102;1601	All

# HLA-B\*27 Single Box 1.0 Typing Kit components

• HLA-B\*27 Single Box typing plates\* (88 typings)

4 plates (22 typings each) (Store at -30 to -15 °C)

PCR Master Mix (With Taq DNA Polymerase)

4 X 80 µl (Store at -30 °C to-15°C)

Plate sealers

12 sealer capsules

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#### **PCR Master Mix Components**

#### **Nucleotides**

Final concentration of each dNTP: 600 µM

#### PCR Buffer

Final concentration: 3,3x NH<sub>4</sub> Buffer; 2,0 mM MgCl<sub>2</sub> and 0,4

U/µl Taq DNA polymerase, pH 8.3.

Glycerol

Final concentration: 16,6%

Cresol Red

Final concentration: 300µg/ml

<sup>\*</sup> With dried specific primers pares.

# PCR amplification protocol

# Reagents

- DNA Sample (100-200 ng/μl)
- PCR Master Mix
- ddH<sub>2</sub>O (not supplied)

#### **DNA Extraction**

For SSP typing highly pure DNA is needed. We recommend isolation of DNA using any extraction kit with CE marking, which guarantees an OD ratio 260/280 higher than 1.6 and a 100ng - 200 ng/µl DNA concentration.

Alternatively, the DNA can be extracted using trimethyammoium-bromide salts (DTAB/CTAB) or by salting out, dissolving it in TE Buffer. The same OD and concentration values should be assured. DO NOT USE HEPARINISED BLOOD WITH THIS METHOD

# **PCR Amplification**

- 1. Spin briefly the DNA and Master Mix tubes.
- 2. Add:
  - 80 µl of PCR Master Mix,
  - 160 µl of ddH₂O

to a 0,7 ml or 1,5 ml tube.

- 3. Vortex the tube vigorously for 15s.
- Load 9 μI of the mix into each tube of the strip (1 primers pair).
- 5. Load 1  $\mu$ I of DNA sample (conc. 100-200 ng /  $\mu$ I) in each well (except to the control wells).

6. Close the typing plate with a strip cap ( sealer capsule) and put it in a 96 well thermo cycler.

# **PCR Cycling Parameters**

Step	Temperature	time	Cycle
Denaturation	96 °C	1 min	1
Denaturation Annealing Extension	96 °C 70 °C 72 °C	25 sec 45 sec 30 sec	5
Denaturation Annealing Extension	96 °C 65 °C 72 °C	25 sec 45 sec 30 sec	21
Denaturation Annealing Extension	96 °C 55 °C 72 °C	25 sec 1 min 2 min	4
Extension	72 °C	10 min	1
Keep (optional)	4 °C	Infinite	1

- 7. Keep the strip at 2 to 8 °C after the PCR have finished.
- Detect PCR products running a 2% agarose gel electrophoresis.

# Gel Electrophoresis protocol

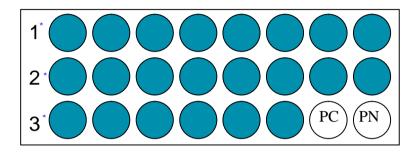
#### PREPARING 2% AGAROSE GEL

- Dissolve 4 grams of electrophoresis grade agarose powder in 200 ml of 1X TAE buffer.
- 2. Melt the agarose powder completely in a microwave oven.
- Cool the heated agarose gel to ~50°C.
- Add at least 10 μl of ethidium bromide<sup>++</sup> (10 mg/ml) or Sybr Safe<sup>TM</sup> (100000 x concentrated) to the heated agarose. Stir until it is thoroughly incorporated.
- 5. On a balanced surface, set up a gel strip with **96 wells**.
- 6. Cast a 5 mm thick gel on the strip.
- 7. Allow the gel to settle.
- \*\* Caution, this reagent is a strong mutagenic agent (read carefully its MSDS before using it).

# **GEL ELECTROPHORESIS**

- 1. Submerge the gel in 1X TAE buffer in a electrophoresis tank.
- 2. Gently remove the caps to avoid splashing of PCR products.
- 3. Load 10 µl into each well on the gel.
- Connect the electric leads and turn on the power supply (115V).
   Electrophoresis for ~ 20 minutes, or until 2/3 of the lane.
- 5. Transfer the gel to a UV transilluminator, document the result by photography.
- 6. Use the **Result Interpretation Sheet** (1) to interpret results.

# HLA-B\*27 Single Box 1.0 plate



\* Numbers may differ fro plate to plate: The no1 may be 4, 7, 10 The no 2 may be 5, 8, 11 the no 3 may be 6, 9, 12

# HLA-B\*27 SINGLE BOX 1.0 plate Identification

Position	Product
Lines 1,2 and 3 (except controls)	B*27
PC	Positive control
NC	Negative control

# **Results Interpretation sheet (1/1)**

Position	HLA-B 27	Specific band	Control band**
Lines 1,2	B*27 positive	150	790 pb
and 3 (except controls)	B*27 negative		<b>790 pb</b>
PC	Positive control	150	
NC	Negative control		790 pb

The PCR reaction is only valid in the presence of control band or, in some cases, in the presence of the specific band.

In the absence of the control band, please repeat the typing.

<sup>\*\*</sup>Control primer pares match with non-allelic sequences. The internal positive control primer pairs amplify segments of the HLA-DRB1 gene, giving rise to 790 base pair fragment. In the presence of the specific band amplification the control band intensity often decreases.

# **Troubleshooting Guide**

PROBLEMS	POSSIBLE CAUSES	SUGGESTIONS
The control and specific		Check DNA quality and concentration
	Concentration of DNA sample is too low.	Re-extract the DNA sample or try not add water into the PCR Mix
bands are weak.		Repeat typing with a good quality DNA sample
	DNA polymerase inhibitors in the	Re-purify the sample DNA
	DNA sample	Repeat typing with a good quality DNA sample
	DNA polymerase inhibitors in the	Re-purify the sample DNA
Missing internal control	DNA sample.	Repeat typing with a good quality DNA sample
bands in one or several lanes.		Check the plate sealing
	Dried PCR amplification products	Repeat the typing using a PCR MicroMat and/or overlay the PCR reaction mix with mineral oil
False negative of a specific band while the internal	Degradation of DNA comple	Re-extract the DNA sample with fresh material
control appears normal	Degradation of DNA sample	Repeat typing with a good quality DNA sample
		Check DNA quality and concentration
	Excess of template DNA	Dissolve the DNA sample in <sub>dd</sub> H2O in order to have the proper concentration
		Repeat typing with a good quality DNA sample
More than two specific alleles are detected/		Clean the working area
Ambiguous results	Contamination with previously	Work in separated pre-PCR and post-PCR rooms
	amplified PCR products or with other DNA samples during the DNA	Keep different lab coats in pre-PCR and post- PCR rooms
	extraction or PCR preparation steps	Change protective gloves frequently
		Repeat typing with a good quality DNA sample
	Demonstration of DNA counts	Re-extract the DNA sample with fresh material
Blurred bands	Degradation of DNA sample	Repeat typing with a good quality DNA sample
		Check DNA quality and concentration
	Excess of template DNA	Dissolve the DNA sample in <sub>dd</sub> H2O in order to have the proper concentration
		Repeat typing with a good quality DNA sample
	Electrophoresis Buffer Problems: wrong buffer or older buffer	Use a fresh recommended buffer

# **Precautions and Warnings**

PCR amplification allows the amplification of small quantities of sample DNA in an exponential way. However, this is also true for foreign DNA, which can contaminate our PCR method. Consequently, special laboratory practices are necessary in order to avoid false positive amplifications. Bellow is listed Genebox recommendations to circumvent contaminations:

- Work in separated pre-PCR and post-PCR rooms.
- Laboratory workflow must be unidirectional, from pre-PCR to post-PCR area.
- Specific equipment for each working area must be used (sample preparation, amplification and preamplification).
- All equipment used in post-PCR should not leave this area.
- Use dedicated micropipettes, gloves and lab coats in each area.
- Use non talcum powder gloves (since talcum could inhibit the PCR reaction).
- Use filter tips in order to avoid cross contamination.
- Check regularly micropipettes, in order to ensure that they are accurate within 5 % of fixed volume.
- Use different micropipettes depending on the volume we wish to load.
- Check regularly thermocyclers, in order to ensure that they are accurate within 1% of fixed temperature.
- Open and close reagent vials carefully. After use, close vials and store at indicated temperatures.
- Do not use a kit after its expiration date.
- packaging material included within the kit is resistant to the indicated storage conditions. Storage at different conditions can cause breakage of the material, and possible contamination of the kit reagents.
- plastic material included within the kit is resistant under normal conditions of use. Use of plastic material in extreme conditions may cause its breakage, and therefore, the impossibility to use the kit.
- check suitability of DNA quantity and quality before use the kit.

#### General instructions for laboratory safety:

- do not eat, drink or smoke in laboratory work areas
- wear disposable gloves
- wear clean lab coats and eye protection
- wash hands thoroughly after handling specimens and test reagents
- clean the working area before and after kit handling.

#### **Technical Guide**

# 1. DNA Quality and Concentration

For optimal results with the HLA-B 27 Single Box 1.0 Typing Kit<sup>™</sup> the quality of DNA is critical. Good quality DNA means an OD ratio 260/280 higher than 1.6 and the major portion of DNA should run higher than 9.4 kb on an agarose qel. Different quality and concentration values require DNA re-extraction.

The quantity of DNA should be 100ng – 200ng/µl. Excess of DNA can cause unspecific amplification.

We recommend any DNA extraction kit which has CE marking, in order to obtain this highly DNA purity

#### 2. Tag Polymerase

HLA-B\*27 Single Box 1.0 Typing Kit<sup>TM</sup> kits have been intensively tested with the Taq DNA Reagente 5 (Reagente 5, Lisbon, Portugal).

#### 3. PCR Master Mix

For optimal results with the HLA-B\*27 Single Box 1.0 Typing Kit<sup>™</sup> the use of the master mix supplied is obligatory.

# 4. Amplification Procedure

At the end of PCR, examine the degree of evaporation and condensation of PCR reaction mixture. If there is more than 20% volume loss do not validate the results. In order to prevent this you should overlay the PCR reaction mixture with mineral oil or use a MicroMat. It is also a good practice to maintain QC records on the heating lid.

If the temperature of the heating lid is not high enough, it will cause condensation problems on the lid.

# 5. Thermal Cycler

We recommend the use of any thermocycler with the following characteristics:

- heating rate up to 2.5°C/sec; cooling rate up to 1.5°C/sec; temperature range 4-100°C; temperature uniformity  $\pm 0.5$ °C; heated lid up to 100°C.

# 6. Validity

As specified in the package

If your problems persist, do not hesitate to contact our technical support

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#### Guarantee

geneBOX - R&D Diagnostic Tests guarantees that the primers in HLA-B 27 single box typing kit have the specificities given in the Results Interpretations Sheet/Tables of the product insert.

# 1. Typing strip

When stored at -20°C, the dried primers are stable for 12 to 19 months from the date of manufacture (see lot validity in the package).

When stored at 4°C, the dried primers are stable for 12 months from the date of manufacture (see lot validity in the package).

At room temperature, the dried primers are stable for 3 to 4 weeks from the date of the reception.

When the sealer is removed the dried primers steal be stable for 2 days, maximum, in dried conditions.

#### 2. PCR Master Mix

When stored at -20°C, the PCR Master Mix is stable for 18 months from the date of manufacture (see lot validity in the package).

When stored at 4°C, the Master mix is stable for 15 days from the date of the reception.

At room temperature, the master mix steal be stable for 3 days from the date of the reception.

The master mix should not be left or stored with the cap open.

#### 3. DNA

Using extracted DNA from salting out or any kit procedure the samples should be stored at 4°C or -20°C. If you chose to freeze the samples you must avoid repeated cycles of heating/freezing, in order to preserve your sample stability.

The DNA samples stored in  $dH_2O$  are stable for 2 to 4 weeks (at 4°C) or 24 months (at -20°C).

The DNA samples stored in buffer are stable for 12 months (at 4°C) or 5 years (at -20°C).

# Warranty

geneBOX - R&D Diagnostic Tests warrants its products to the client against defects in materials and contents under normal application. The company products under this warranty shall be replaced, at no charge, to the damaged client.

This warranty applies only to products that have been handled and stored in accordance with its recommendations/specifications.

The claims must be posted directly to geneBOX in writing and must be accompanied by a copy of the purchaser's invoice.

This product may not be reformulated, repacked or resold in any form without geneBOX - R&D Diagnostic Tests consent.

# **Declaration of conformity**

Product Name: HLA-B\*27 Single Box

Product Number: GB.10.06

**Intended use:** HLA-B\*27 allele testing.

Manufacturer: geneBOX - R&D Diagnostic Tests,

Biocant – centro de inovação em biotecnologia

núcleo 4, lote 3

3060-197 Cantanhede,

Portugal

We, geneBOX - R&D Diagnostic Tests, hereby declare that this product, to which this declaration of conformity relates, is in conformity with the following standards and other normative documents ISO 9001:2008 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices as transposed into the national laws of the Member States of the European Union.

The technical file of the product is maintained at geneBOX - R&D Diagnostic Tests, Biocant Park, Parque tecnológico de Cantanhede, 3060-197 Cantanhede, Portugal.

Sandra Balseiro Technical Director

# Material Safety Data Sheet (MSDS) (1/3)

# geneBOX - R&D Diagnostic Tests<sup>™</sup> PCR-SSP Kits

# geneBOX <sup>™</sup> PCR-SSP typing products

This Material Safety Data Sheet (MSDS) applies to all geneBOX - R&D Diagnostic Tests SSP™ typing kits

#### 1. Chemical products and company identification

Date of Issue: May 2010

Product group: geneBOX<sup>™</sup> PCR-SSP Typing Products Manufacturer: geneBOX - R&D Diagnostic Tests,

biocant - centro de inovação em biotecnologia

núcleo 4, lote 3

3060-197 cantanhede, portugal

tel/fax: +351 231 410 946/ +351 231 410 947

e-mail: info@genebox.com

#### 2. Composition and reagents information

 Component
 Chemical
 Common Name

 Plate
 Deoxyribonucleic acid
 Oligonucleotide

 Cresol Red
 PCR Master Mix
 Deoxyribonucleotides
 Nucleotides

NH<sub>4</sub> Buffer Magnesium chloride MgCl2

Cresol Red

Glycerol Glycerine

#### 3. Physic-chemical properties:

 Components
 Appearance
 Colour
 Odour

 Plate
 dried, in plate wells
 Red
 none

 Master Mix
 liquid
 Pink/red
 none

# 4. Toxicological information

Chemical Toxicities

Glycerol LD50= oral 4090 mg/kg (mouse) LD50= oral 12600 mg/kg (rat) LD50= oral 1480 mg/kg (human)

#### 5. Stability and reactivity

Conditions to avoid: Heat and moisture.

**Incompatibilities:** Strong oxidizing agents, strong bases.

# Material Safety Data Sheet (MSDS) (2/3)

#### 6. Personal protection.

**Hand protection:** Wear appropriate chemically resistant gloves. **Eye protection:** Chemical safety goggles are recommended.

**Skin protection:** Wear laboratory coat.

#### 7. Handling and storage

Handling: Avoid substance contact.

Storage: Protect from light. Store at temperature indicated on package.

Package Damage: reject damaged components.

#### 8. Hazards

Master Mix Components: may be harmful by inhalation, ingestion or skin absorption. May cause eye and skin irritation. Material is irritating to mucous membranes and upper respiratory tract. Ingestion of large amounts can cause stomach pains, vomiting or diarrhoea.

#### 9. First aid measures

**In case of eye contact:** Immediately flush eyes with large amounts of water for at least 15 minutes. Call a physician.

In case of skin contact: Immediately wash skin with soap and large amounts of water. Wash contaminated clothing before re-use.

**In case of ingestion:** Wash out mouth with water provided person is conscious. Call a physician if needed.

**In case of inhalation:** remove to fresh air, if not breathing give artificial respiration. If breathing difficult, give oxygen. Call a physician.

#### 10. Fire fighting measures

**Extinguishing media:** Water, carbon dioxide, dry chemical powder or appropriate foam.

Extinguishing media NOT to use: None are known.

**Special exposure hazards:** May emit toxic fumes of carbon dioxide, carbon monoxide, nitrogen, phosphorus, hydrogen chloride, and hydrogen gas under fire conditions.

**Special fire-fighting equipment:** When large amounts of substances are released work only with self-contained breathing apparatus and protective clothing to prevent contact with skin and eyes.

#### 11. Accidental release measures

**Personal Precautions:** Avoid substance contact. No further requirements. **Cleaning Method:** Clean up affected area. No further requirements.

# Material Safety Data Sheet (MSDS) (2/3)

#### 12. Ecological information

No data available.

#### 13. Waste disposal information

Waste disposes in accordance with all applicable regulations (the disposals should be incinerated).

#### 14. Transport information

During transportation the temperature could not exceed 25°C. Transportation should not exceed 3 days.

#### 15. Other information

The above information is based on our current level of knowledge, but does not purport to be all-inclusive and shall be used only as a guide. <code>geneBOX - R&D Diagnostic Tests</code> shall not be held liable for any damage resulting from handling or from contact with the above products.

If your problems persist, do not hesitate to contact our technical support

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DESENVOLVIMENTO E PRODUÇÃO DE TESTES DE DIAGNÓSTICO

biocant – centro de inovação em biotecnologia núcleo 4, lote 3 3060-197 cantanhede portugal

tel + 351 231 410 946 fax + 351 231 410 947 e-mail info@genebox.com www.genebox.com